



## Green synthesis of silver nanoparticles using soymida febrifuga: optimization, characterization and for dye degradation in photocatalytic activity via green way

Varsha. S. Nandeshwar<sup>1,a</sup>, Surekha. A. Kalkar<sup>1,b</sup>, Pratibha. S. Agrawal<sup>2,c</sup>

<sup>1a</sup>Research scholar, Department of Botany, Institute of Science, RTMNU, Nagpur (M.H) 440008, India.

<sup>1a</sup> Professor, Department of Botany, Institute of Science, RTMNU, Nagpur (M.H) 440008, India.

<sup>2c</sup> Professor, Department of Applied Chemistry, LIT, RTMNU, Nagpur (M.H.) 440033, India.

Email - <sup>1</sup>varshanandeshwar26@gmail.com

**Abstract:** Traditional synthetic techniques for silver nanoparticles synthesis involve toxic chemicals that are harmful to humans as well as the environment. The green chemistry method for nanoparticle synthesis is rapid, eco-friendly, and less toxic as compared to the traditional methods. In the present research, we synthesized silver nanoparticles employing a green chemistry approach from *Soymida febrifuga* bark extract. The optimized *Soymida febrifuga* silver nanoparticles (SFAGNPs) had a mean particle size of  $187.87 \pm 4.89$  nm with a narrow size distribution of  $0.226 \pm 0.009$  and surface charge  $-34 \pm 3.12$  mV, respectively. The physicochemical characterization of optimized SFAGNPs was done by Fourier transform infrared spectroscopy (FTIR). These SFAGNPs were employed for the treatment of wastewater and dye degradation and so on.

**Key Words:** silver nanoparticles; *soymida febrifuga*; green chemistry; dye degradation

### 1. INTRODUCTION:

Reported by S.S. Patil (2016) in their book "In the last few decades, anthropogenic activities and rapid industrialization have resulted into severe water pollution and thereby posing a serious threat towards human and ecological environment. In particular, organic and inorganic dyes with more than  $7 \times 10^5$  tons annuals productions are the major toxic water pollutants. A large portion of it means more than 15% toxic and carcinogenic dyes is being discharged into water annually. However, only 47% of the synthetic dyes are biodegradable and generally require a significant attention due to their recalcitrance and hazardous characteristics. Even at low concentration, occurrence of dye affects the aquatic photosynthetic ecosystem and can be detrimental for aquatic lives as their growth can be inhibited by interfered sunlight in water bodies by organic dye molecules". Therefore green chemistry is the utilization of chemistry principles to minimize the usage of poisonous chemicals, minimal unwanted residues, which might be injurious to human health as well as to the atmosphere(1,2). A combination of green chemistry and nanotechnology is a versatile approach and has extended its consideration in the last few years (3,4). The green synthesis of nanoparticle (NPs), which is ecological and cost-effective and employs stabilizing agents attained from plant extracts and other natural resources for the fabrication of NPs without the usage of harmful compounds, also promotes the sustainable use of NPs (5,6). In the recent era, metallic NPs have been extensively used for various biomedical applications such as diagnosis, drug-delivery systems (DDs), and tissue engineering because of their exceptional physicochemical properties.....(7,8). Among numerous metallic NPs, silver nanoparticles (AgNPs) have gained much attention for various applications, including imaging contrast agents, sensors, and antimicrobial agents owing to their better stability, electrical conductivity, and antimicrobial activity (9,10).

Also, AgNPs have shown their potential for the management of cancer. Many in vitro studies using AgNPs have proved their action as active anticancer agents against various cancer cells, including lung carcinoma, breast cancer, and human cervical cancer cells..... (11,12). Generally, AgNPs are synthesized via several chemical and physical approaches. The physical methods consist of ball milling, flame sonication, radiation, electric arc discharge, pyrolysis, etc.....(13,14). These physical methods often require expensive instruments, higher pressure, and more energy consumption (9,11,15). Though, the chemical approaches frequently need expensive metal salts and harmful solvents. Moreover, different stabilizers are also essential to avoid the aggregation of NPs to make them physiologically



compatible (16,17). In this context, green synthesis of AgNPs by plant extracts as a substitute method for the traditional physical and chemical NPs preparation methods.....(18,19).

*Soymida febrifuga* / *Swietenia febrifuga* (Roxb.) A. Juss. (Family-Meliaceae), generally bark used in the treatment of diarrhea, dysentery and fever and also as a general tonic, decoction used in gargles, vaginal infections, rheumatism swellings and as enemata. The deep red bark is very astringent and bitter. It is used in the treatment of diarrhea and dysentery, and is also often used as a febrifuge instead of quinine (*Cinchona* spp.) by local people. It is said to be an efficient remedy for the dangerous jungle fever of India when quinine produces no effect. It has also been employed successfully against typhus fever and has proven to be effective in treating bad cases of gangrene. But this *Soymida* plant not used for water treatment and dye degradation till now (20,21). Although conventional weed control methods are frequently used to manage this problematic plant, exploring its utility potential as an innovative management strategy is gaining attention nowadays (22,23). The plant is well-known for its primary and secondary metabolites, such as proteins, carbohydrates, tannins, volatile oils; especially lupeol, sitosterol, methyl angolensate, deoxyandirobin and 2 tetranortriterpenoids with a modified furan ring from the bark that contribute towards its allelopathic capacity, cytotoxic activity, and allergenic reactions (24,25). However, studies have reported that biochemical from *Parthenium hysterophorus* also have medicinal values such as antioxidant, anti-inflammatory, antibacterial, antifungal, and anticancer activities (26,27). There have been efforts in preparing NPs from the bark extracts of *Soymida febrifuga*, and further work was suggested to develop the value-added products of biomedical significance (28,29). But no study is performed yet for the optimized synthesis of AgNPs from this plant and also detailed therapeutic investigation of the so-formulated NPs has been investigated. The main objective of this study was the synthesis, optimization, and characterization of SFAgNPs from *Soymida febrifuga*, as well as the detailed study of their therapeutic activity, which to our best knowledge has not been done so far. For attaining this purpose in this study, we fabricated silver NPs using bark of *Soymida febrifuga* extract using a green chemistry approach. Synthesized SFAgNPs were characterized by UV-Vis spectroscopy, and the surface morphology of developed NPs was characterized by scanning electron microscopy (SEM) respectively. Moreover, the solid-state characterization had done by Fourier transform infrared spectroscopy (FTIR).

## 2. MATERIALS AND METHODS:

### Preparation of Bark Extract

Fresh bark of *Soymida febrifuga* were collected from our campus Institute of Science, Nagpur (M.H.). Bark were thoroughly rinsed with tap water and then with double-distilled water. The bark were shade dried for 2-3 weeks. Uniform powder was made by passing the dry bark through a 50 mm sieve. To prepare bark extract, 10 g of bark powder was mixed with 100 mL distilled water followed by boiling at 80 °C for 30 min. Then, the extract was filtered by Whatman filter paper (No.1) and stored at 4 °C before being used to biosynthesize silver nanoparticles.



Figure 1. Bark of *Soymida febrifuga* plant.

### Phytoreduction of *Soymida febrifuga* Silver Nanoparticles (SFAgNPs)

For silver nanoparticles synthesis, 1 mL of the *Soymida febrifuga* bark extract was added dropwise into 9 mL of freshly prepared 1 mM AgNO<sub>3</sub> solution with continuous stirring at room temperature for 30 min. The synthesis of nanoparticles was confirmed by reddish-brown color. After that solution was kept in the dark for a further 24 h allowing the complete reduction of the silver nitrate solution. UV-Vis spectrophotometry also confirmed the synthesis of silver nanoparticles. Lastly, SFAgNPs were collected by centrifugation at 10,000 × g rpm for 15 min by using optimized conditions (30,31). Different concentrations of AgNO<sub>3</sub> i.e., 0.5 mM, 1 mM, 2 mM, 3 mM, and 4 mM and bark extract i.e., 1 mL, 2 mL, 3 mL, and 4 mL and different reaction time i.e., 30 min, 1 h, 4 h, 6 h, 12 h, and 24 h were employed to standardize the optimum conditions.

**Visual Inspection:**

The color change after mixing silver nitrate with the plant extract in a suitable ratio is the first indicator of the formation of silver nanoparticles (32,33). The color change was monitored at various time intervals after reducing silver nitrate with *Soymida febrifuga* bark extract for checking the synthesis of SFAgNPs.

**UV-Visible Spectroscopy:**

The synthesis of silver nanoparticles was investigated by checking the absorbance of the diluted solution of silver nanoparticles suspension at a wavelength of 300–800 nm by a UV-Vis spectrophotometer. Silver nanoparticles exhibit strong electromagnetic wave absorption in the visible range owing to the surface plasmon resonance (SPR).

**Scanning Electron Microscopy (SEM):**

Morphology of the synthesized SFAgNPs was investigated by scanning electron microscope. For analysis, SFAgNPs were placed on glass slide following coating them with a thin layer of gold-palladium blend once mounted them on a carbon grid.

**Fourier Transform Infrared Spectroscopy (FTIR):**

FTIR analysis was used to scan the synthesized silver nanoparticles to obtain the capped functional groups on the nanoparticles. FTIR analysis of the *Soymida febrifuga* bark extract and synthesized nanoparticles was carried out using the KBr pellet method by the FTIR apparatus in transmittance mode from 400–4000  $\text{cm}^{-1}$  at 4  $\text{cm}^{-1}$ .

**Treatment of Dye Degradation with under the photo catalytic activity Different dyes by SFAgNPs:**

Synthetic dyes are extensively used in textile dyeing, paper printing, color imaging, pharmacy, food, cosmetics, leather industries and laboratory and so on. These dyes are poisonous and recalcitrant in nature due to their complex chemical structures. For better elimination of dye wastewater, efficient photocatalysts should be synthesized. Degradation of Acid orange 7 (AO7), often known as Orange II, Azo black (Chlorazole Black), Indigocarmine, Tartrazine, and Alizarine Red (10Tml/L) under UV light and visible irradiation by SFAgNPs. Dye degradation by catalysis becoming a key agent in self-care products, i.e., disinfectant or bleaching agents and broadly supplied in water or wastewaters.

**3. Results and Discussion :****Visual Inspection**

The change of color from dark brown to dark greenish in color, thereby visually confirmed the production of silver nanoparticles. Color change was observed within a half-hour of exposure (Figure 2). The change in solution color is due to the surface plasmon resonance (SPR) displayed by the bio-reduced AgNPs (34,35). These results are in accordance with previously reported work (36,37)



Figure 2. Formation of Silver NPs from *Soymida febrifuga*





### UV-Visible Spectroscopy

The ultraviolet-visible absorption spectrum of the synthesized SFAgNPs dispersion was examined between 300–800 nm through a UV-visible spectrophotometer. The spectrum of the SFAgNPs dispersion showed a single surface plasmon resonance absorption band with a maximum of approximately 432 nm (Figure 3). As the symmetry of NPs increased, the number of SPR peaks decreased, and spherical SFAgNPs exhibited only one peak, while triangular and disc shape NPs showed two or more peaks. As the particle size increases, the SPR of the absorption spectrum of metal nanoparticles typically shifts to longer wavelengths (38,39). The effect of varying concentrations of silver nitrate and leaf extract and reaction time on the formation of SFAgNPs was also studied by periodically checking the absorbance of the reaction mixture through UV-Vis spectroscopy at 300–800 nm.

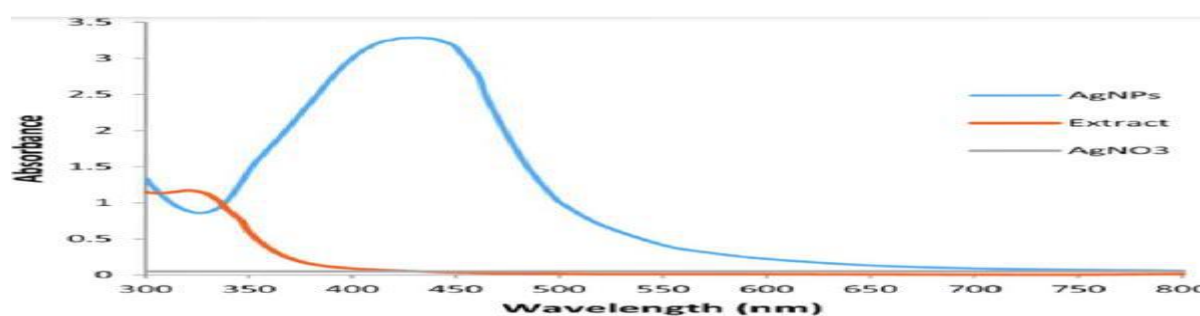


Figure 3. UV-Visible spectroscopy of AgNO<sub>3</sub> Soymida febrifuga plant extract and silver NPs of Soymida febrifuga plant.

### Effect of Reaction Time:

The optimal time for the synthesis of SFAgNPs was examined through the UV-Vis spectra of the reaction mixture at various time intervals. As the reaction time increases, the redshift of the absorption peak designates an increase in the size of the NPs (Figure 2A). The increase in the intensity of the absorption peak over time is due to the increase in the number of absorbed metallic NPs (40,41). The synthesis of AgNPs from *Solanum trilobatum* extract was also studied by Vanaja et al., who observed a sharp narrow peak at 420 nm after 20 min of reaction indicating the synthesis of AgNPs, which was shifted to 440 nm after 20 min. With an increase of reaction time, the AgNPs production went on increasing, and the maximum absorption peak denoted the highest yield. The completion of the reaction occurred within 4h, which was also visualized by precipitate formation at the bottom (42,43).

### Effect of Silver Nitrate Concentration:

The optimal concentration for the synthesis of SFAgNPs was found by varying concentrations of silver nitrate. The optimal concentration for NPs synthesis was found to be 1 mM (Figure T2B). Usually, by increasing the AgNO<sub>3</sub> concentration, the fabrication of AgNPs is increased, and the color of the solution is enhanced because the accumulation of silver ions takes place, and larger NPs are attained (44,45). As the concentration of silver nitrate increases, the plasmon resonance (SPR) band becomes wider (46,47). Similarly, 1mM silver nitrate was found to be optimal for the synthesis of silver nanoparticles from *Garcinia mangostana* extract (15)(48).

### SEM Investigation:

The SEM images of prepared SFAgNPs showed that these nanoparticles are spherical. Some small NPs aggregated into larger particles, which is due to the existence of secondary metabolites occurring in the *Soymida febrifuga* bark extract (Fig.4). The SEM figure showed a wide distribution of prepared NPs ranging from 10 to 130 nm in size.

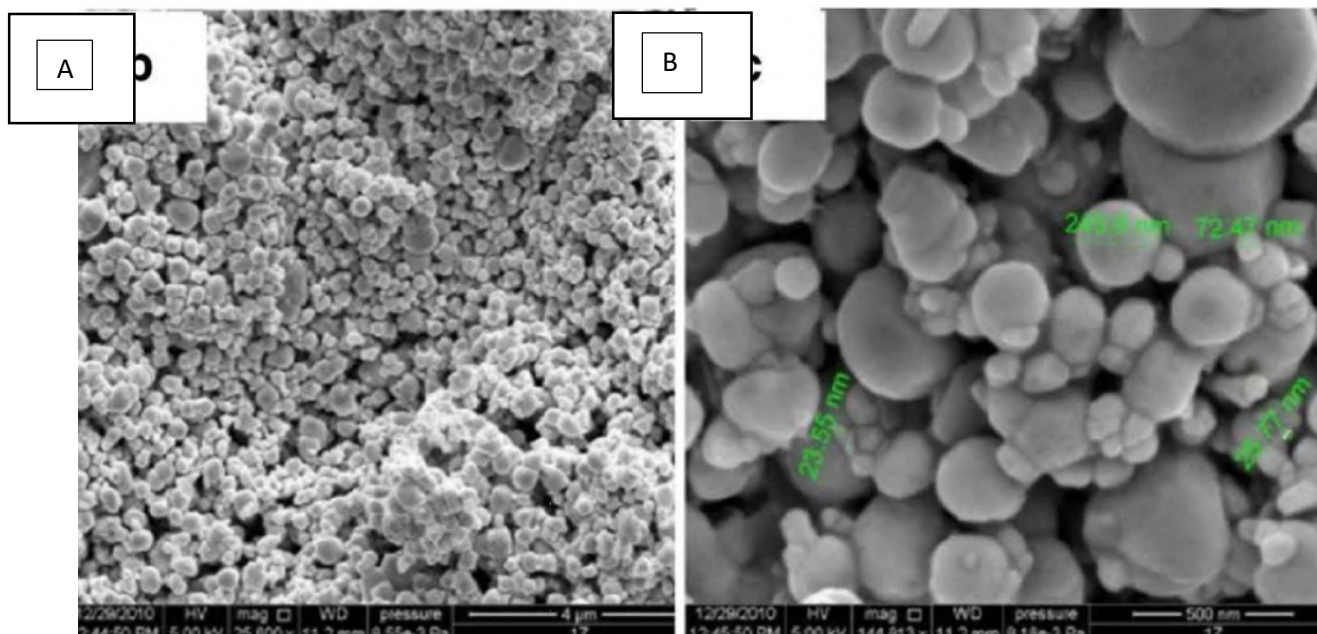


Figure 4. SEM pictures A and B of biosynthesized SFAgNPs.

### FTIR Analysis

FTIR characterized the bark extract of *Soymida febrifuga* and the resultant biogenic SFAgNPs. The FTIR spectra for the bark extract and SFAgNPs are shown in FTIR graph. The characteristic absorbance bands for *Soymida febrifuga* and SFAgNPs can be seen at  $1631\text{cm}^{-1}$ ,  $1588\text{cm}^{-1}$ ,  $1384\text{cm}^{-1}$ ,  $1400\text{cm}^{-1}$ ,  $1126\text{cm}^{-1}$ ,  $759\text{cm}^{-1}$  at the wavelength range of  $400\text{--}4000\text{cm}^{-1}$

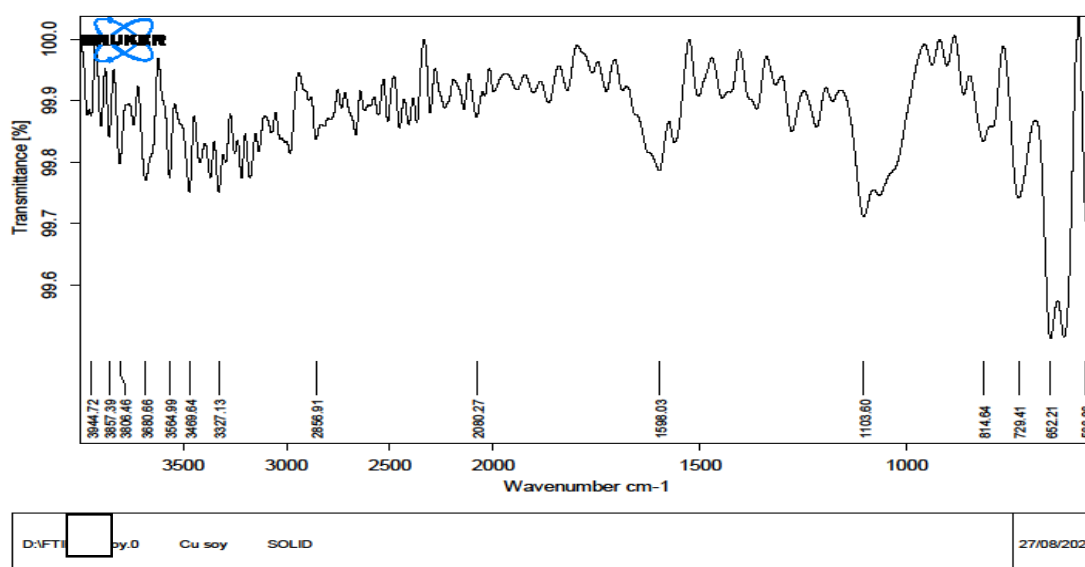


Figure 5. FTIR spectra of Silver Nanoparticles of *Soymida Febrifuga* bark

Which proved the presence of major functional groups of plant extract in the prepared NPs confirming capping and reduction of SFAgNPs with the plant extract.



### Removal of Dyes by SFAgNPs:

The ultra-fast catalytic reduction of dye pollutants using biosynthesized AgNPs can be discussed based on surface adsorption behavior of dyes. Generally, the biological molecules or surfactants are used for capping of nanoparticles in chemical reduction methods, which makes an insulating layer around the nanoparticles and affect the diffusion of dye molecules to adsorb efficiently on surface of metal particles. However, the biomolecules can easily be separated by simple washings with water and thus does not restrict the adsorption of dye molecules on metal surfaces, which makes them efficient catalysts. AgNPs catalyzed reduction of pollutants can also be explained by electrochemical means, in which the AgNPs behave as an electron relay for oxidant and reductant

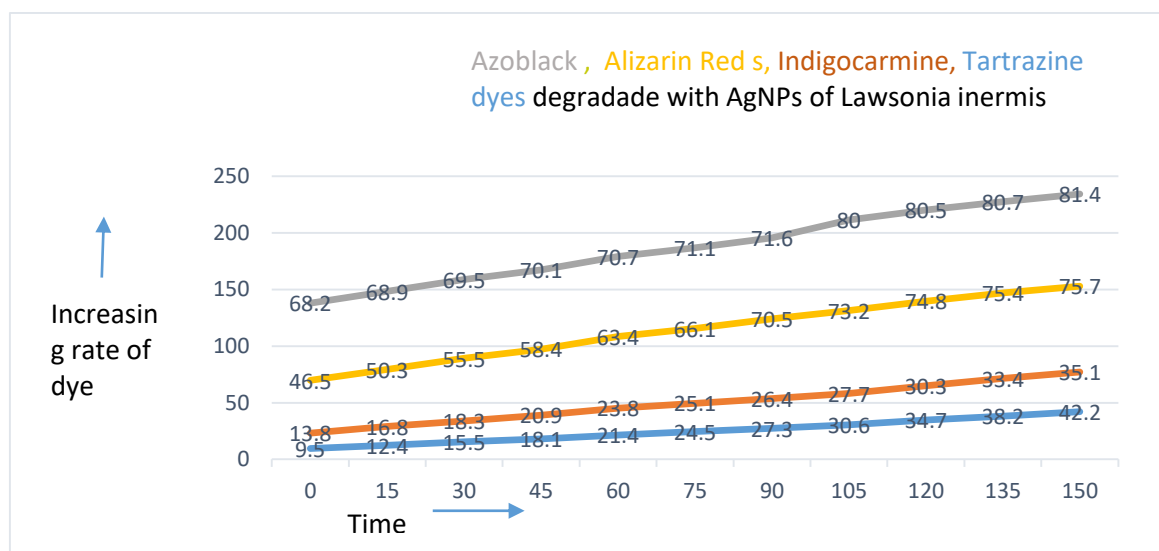


Figure 7: Degradation of Azoblack, Alizarine, Indigocarmine and Tartrazine dyes with AgNPs of *Soymida Febrifuga* (Mansrohini) plant with increasing time with photo catalytic activity.

### 4. CONCLUSIONS:

The green chemistry approach was employed for the rapid and effective synthesis of silver nanoparticles using *Soymida febrifuga* bark extract without any harmful side effects. These synthesized SFAgNPs were successfully optimized by UV-Vis spectroscopy by using different ratios of silver nitrate and bark extract, and reaction time. Phytochemical screening of the *Soymida febrifuga* bark extract was done to confirm which functional groups are present in the extract which was further confirmed by FTIR analysis when the detected function groups (Flavonoids, terpenoids, steroids, amides, alkaloids, and tannins) caused the reduction and stabilization of the synthesized NPs. The size and morphology of the SFAgNPs were examined by SEM confirming the spherical-shaped nanoparticles with size ranging from 20–25 nm. The synthesized AgNPs exhibited excellent role as wastewater disinfectant and dye degradation under the photocatalytic activity and it showed to be decreased by 60-80%, confirming its role in disinfecting wastewater.

### REFERENCES:

1. Khan ZUH, Khan A, Chen Y, Shah NS, Muhammad N, Khan AU, et al. Biomedical applications of green synthesized Nobel metal nanoparticles. *J Photochem Photobiol B Biol.* 2017 Aug 1;173:150–64.
2. Rasheed T, Bilal M, Iqbal HMN, Li C. Green biosynthesis of silver nanoparticles using leaves extract of *Artemisia vulgaris* and their potential biomedical applications. *Colloids Surfaces B Biointerfaces.* 2017 Oct 1;158:408–15.
3. Shubhashree KR, Reddy R, Gangula AK, Nagananda GS, Badiya PK, Ramamurthy SS, et al. Green synthesis of copper nanoparticles using aqueous extracts from *Hyptis suaveolens* (L.). *Mater Chem Phys.* 2022 Mar 15;280.
4. Parveen M, Kumar A, Khan MS, Rehman R, Furkan M, Khan RH, et al. Comparative study of biogenically synthesized silver and gold nanoparticles of *Acacia auriculiformis* leaves and their efficacy against Alzheimer's and Parkinson's disease. *Int J Biol Macromol.* 2022 Apr 1;203:292–301.
5. Saratale RG, Karuppusamy I, Saratale GD, Pugazhendhi A, Kumar G, Park Y, et al. A comprehensive review on green nanomaterials using biological systems: Recent perception and their future applications. *Colloids Surfaces B Biointerfaces.* 2018 Oct 1;170:20–35.



6. Nelsonjoseph L, Vishnupriya B, Ramasamy, Bharathi D, Thangabalu S, Rehna P. Synthesis and characterization of silver nanoparticles using *Acremonium borodinense* and their anti-bacterial and hemolytic activity. *Biocatal Agric Biotechnol*. 2022 Jan 1;39.
7. Barani D, Benhaoua B, Laouini SE, Bentemam H, Allag N, Berra D, et al. GREEN SYNTHESIS OF ZnO NANOPARTICLES USING PHOENIX DACTYLIFERA . L LEAF EXTRACT: EFFECT OF ZINC ACETATE CONCENTRATION ON THE TYPE OF PRODUCT. 2019;14(3):581–91.
8. Sharmila G, Muthukumaran C, Sandiya K, Santhiya S, Pradeep RS, Kumar NM, et al. Biosynthesis, characterization, and antibacterial activity of zinc oxide nanoparticles derived from *Bauhinia tomentosa* leaf extract. *J Nanostructure Chem* [Internet]. 2018;8(3):293–9. Available from: <https://doi.org/10.1007/s40097-018-0271-8>
9. Abdelghany TM, Al-rajhi AMH, Abboud MA Al. Recent Advances in Green Synthesis of Silver Nanoparticles and Their Applications : About Future Directions . A Review. 2017;
10. Santhoshkumar J, Kumar SV, Rajeshkumar S. Resource-Efficient Technologies Synthesis of zinc oxide nanoparticles using plant leaf extract against urinary tract infection pathogen. *Resour Technol* [Internet]. 2017;3(4):459–65. Available from: <https://doi.org/10.1016/j.reffit.2017.05.001>
11. Thandavamoorthy P, Thiruvengadam D. Innovare Academic Sciences GREEN SYNTHESIS , CHARACTERIZATION , ANTIMICROBIAL AND CYTOTOXIC EFFECTS OF SILVER NANOPARTICLES USING ORIGANUM HERACLEOTICUM L . LEAF EXTRACT. 2015;7(4).
12. Banna A, Banna LS Al, Salem NM, Jaleel GA, Awwad AM. Green synthesis of sulfur nanoparticles using *Rosmarinus officinalis* leaves extract and nematicidal activity against *Meloidogyne javanica*. 2020;6(3):137–43.
13. Awwad AM, Salem NM, Aqarbeh MM, Abdulaziz FM. Green synthesis , characterization of silver sulfide nanoparticles and antibacterial activity evaluation. 2020;6(1):42–8.
14. Arts N, Arts N. ANTIOXIDANT , ANTIMICROBIAL AND SEWAGE TREATMENT OF SYNTHESISED SILVER NANOPARTICLES. 2017;4(9345269943):2350–61.
15. Singh J, Dutta T, Kim KH, Rawat M, Samddar P, Kumar P. “Green” synthesis of metals and their oxide nanoparticles: Applications for environmental remediation. *Journal of Nanobiotechnology*. 2018.
16. Sridhar S. Green Synthesis, characterization and in-vitro antibacterial activity of silver nanoparticles by using *Tinospora cordifolia* leaf extract. 2020;(January 2012).
17. Peter S. GREEN SYNTHESIS OF SILVER NANOPARTICLES USING LEAF EXTRACT OF AYAPANA TRIPLINERVIS AND ITS ANTIBACTERIAL ACTIVITY Sona S. Dev \* , K. U. Vineetha and Sneha Francis Department of Biotechnology, St. Peter’s College, Kolenchery, Kochi - 682311, Kerala, India. 2018;9(9):3897–902.
18. Shiekh RA, Balam SK. Biogenic silver nanoparticles using *Rhinacanthus nasutus* leaf extract : synthesis , spectral analysis , and antimicrobial studies. 2013;3355–64.
19. Kaushal A, Sk S. Adsorption Phenomenon and Its Application in Removal of Lead from Waste Adsorption Phenomenon and Its Application in Removal of Lead from Waste Water : A Review. 2017;(October).
20. Ahmed S, Ikram S. Silver Nanoparticles : One Pot Green Synthesis Using *Terminalia arjuna* Extract for Biological Application Nanomedicine & Nanotechnology. 2015;6(4).
21. Anis S. Green Synthesis of Metal-Based Nanoparticles and Their Applications. 2018.
22. Mohammadi F, Yousefi M, Ghahremanzadeh R. Green Synthesis , Characterization and Antimicrobial Activity of Silver Nanoparticles ( AgNps ) Using Leaves and Stems Extract of Some Plants. 2019;2(4):266–75.
23. Yusof HM, Mohamad R, Zaidan UH, Aini N, Rahman A. Microbial synthesis of zinc oxide nanoparticles and their potential application as an antimicrobial agent and a feed supplement in animal industry : a review. 2019;1–22.
24. Khorrami S, Abdollahi Z, Eshaghi G, Khosravi A. An Improved Method for Fabrication of Ag-GO Nanocomposite with Controlled Anti-Cancer and Anti-bacterial Behavior ; A Comparative Study. *Sci Rep* [Internet]. 2019;(June):1–10. Available from: <http://dx.doi.org/10.1038/s41598-019-45332-7>
25. Nadeem M, Abbasi BH, Younas M, Khan T. Green Chemistry Letters and Reviews A review of the green syntheses and anti-microbial applications of gold nanoparticles. 2017;8253. Available from: <https://doi.org/10.1080/17518253.2017.1349192>
26. Selvam K, Sudhakar C, Govarthanan M, Thiyagarajan P, Sengottaiyan A, Selvankumar T. Eco-friendly biosynthesis and characterization of silver nanoparticles using *Tinospora cordifolia* ( Thunb .) Miers and evaluate its antibacterial , antioxidant potential. *J Radiat Res Appl Sci* [Internet]. 2019;10(1):6–12. Available from: <http://dx.doi.org/10.1016/j.jrras.2016.02.005>
27. Mathiyalagan R, Markus J, Kim Y, Wang C, Singh P. Green synthesis of multifunctional silver and gold





- nanoparticles from the oriental herbal adaptogen : Siberian ginseng. 2016;3131–43.
28. Sundar JS, Christobel RGJ, Selvi NK, Abirami MP, Samuel S. Efficient green synthesis of silver nanoparticles from *Caesalpinia bonducella* seeds and its antibacterial and cytotoxic effects : An in vitro study. 2018;7(11):95–102.
  29. Mishra P, Singh L, Mishra S. Biosynthetic Silver Nanoparticles- Current Trends and Future Scope : An Overview. 2019;14(6):37–43.
  30. Omar AA, Alkelbash HM, Alhasomi YF. Green synthesis of silver nanoparticles using olive pomace extract. 2018;662–9.
  31. Nishanthi R, Malathi S, S JP, Palani P. Materials Science & Engineering C Green synthesis and characterization of bioinspired silver , gold and platinum nanoparticles and evaluation of their synergistic antibacterial activity after combining with different classes of antibiotics. Mater Sci Eng C [Internet]. 2019;96(December 2017):693–707. Available from: <https://doi.org/10.1016/j.msec.2018.11.050>
  32. Fakhari S, Jamzad M, Fard HK. Green Chemistry Letters and Reviews Green synthesis of zinc oxide nanoparticles : a comparison. 2019;8253.
  33. Shukla P. Microbial Nanotechnology for Bioremediation of Industrial Wastewater. 2020;11(November).
  34. Masum MI, Siddiqua MM, Ali KA, Zhang Y. Biogenic Synthesis of Silver Nanoparticles Using *Phyllanthus emblica* Fruit Extract and Its Inhibitory Action Against the Pathogen *Acidovorax oryzae* Strain RS-2 of Rice Bacterial Brown Stripe. 2019;10(April):1–18.
  35. Iravani S, Varma RS. Plant Pollen Grains : A Move Towards Green Drug and Vaccine Delivery Plant Pollen Grains : A Move Towards Green Drug and Vaccine Delivery Systems. Nano-Micro Lett [Internet]. 2021;(December). Available from: <https://doi.org/10.1007/s40820-021-00654-y>
  36. Online IP, Polash SA, Nadaf NY, Rahman A, Shohael AM. Green synthesis of silver nanoparticles ( AgNPs ): Agricultural applications and future vision Department of Biotechnology and Genetic Engineering , Faculty of Biological Sciences ,. 2018;13(2):35–57.
  37. Li JF, Liu YC, Chokkalingam M, Rupa EJ, Mathiyalagan R, Hurh J, et al. Phytosynthesis of silver nanoparticles using rhizome extract of *Alpinia officinarum* and their photocatalytic removal of dye under UV and visible light irradiation. Optik (Stuttg). 2020 Apr 1;208.
  38. Hazra I, Sourav C. Green synthesis of water-dispersible silver nanoparticles at room temperature using green carambola ( star fruit ) extract. 2014;
  39. Krishnadhas L, Santhi R, Annapurani S. Green Synthesis of Silver Nanoparticles from the Leaf Extract of *Volkameria inermis*. 2017;9(8):610–6.
  40. Dudhane AA, Waghmode SR, Dama LB, Vaibhav P. Synthesis and Characterization of Gold Nanoparticles using Plant Extract of *Terminalia arjuna* with Antibacterial Activity. 2019;(June).
  41. Of S, Oxide Z, From N, Of F, By A, Andgreen C, et al. Silver nanoparticles obtained by aqueous or ethanolic aloe Vera extracts: An assessment of the antibacterial activity and mercury removal capability. J Nanomater. 2018;
  42. Arts KKW, Science C, Nashik C. PHYTOCHEMICAL ANALYSIS AND BIOLOGICAL ACTIVITIES OF *SOYMIDA FEBRIFUGA* ( ROXB .) JUSS ( MELIACEAE ): AN OVERVIEW. 2019;6(2):826–34.
  43. Teli MD, Sahoo MR, Pandit P, Tech M. Antibacterial and UV- Protective Cotton fabric made by Herbal Synthesized Silver Nanoparticles. 2017;1310–21.
  44. Nasrollahzadeh M. Efficient catalytic reduction of nitroarenes and organic dyes in water by synthesized Ag / diatomite nanocomposite using *Alocasia macrorrhiza* leaf extract. J Mater Sci Mater Electron [Internet]. 2018;0(0):0. Available from: <http://dx.doi.org/10.1007/s10854-018-9802-9>
  45. Elshamy AM, Rabeh MA, Salem M, Samir A, Muhsinah A Bin, Alsayari A. Antiviral potential of green synthesized silver nanoparticles of *Lampranthus coccineus* and *Malephora lutea*. 2019;
  46. Gandhi N, Sirisha D, Sharma VC. Microwave-Mediated Green Synthesis of Silver Nanoparticles Using *Ficus Elastica* Leaf Extract and Application in Air Pollution Controlling Studies. 2014;4(1):61–72.
  47. Ahmad S, Munir S, Zeb N, Ullah A, Ali J, Bilal M, et al. Green nanotechnology : a review on green synthesis of silver nanoparticles — an ecofriendly approach. 2019;
  48. Lal SS, Nayak PL. Green synthesis of gold nanoparticles using various extract of plants and spices. Int J Sci Innov Discov. 2012;2(3):325–50.