



Microbial Alkaline Protease: A Review

¹Moin Akhtar R. Qazi, ²Vishal R. Parate

Department of Food Technology, University Institute of Chemical Technology (UICET),
Kavayitri Bahinabai Chaudhary North Maharashtra University (KBCNMU), Jalgaon, India

*Email: moinqazi@gmail.com

Abstract: Proteins have for some time been utilized as compound choices to work on the proficiency and financial matters of different modern frameworks and cycles. Basic proteases are proteins having a place with the hydrolases bunch. It performs hydrolysis of peptide obligations of proteins. Proteins are separated into more modest peptides and amino acids by protease chemicals, what capability as biocatalysts. Soluble protease chemicals can be gotten from microorganisms at a sensible cost, and they can create the necessary material constantly and dependably. Alkaline proteases are used in a wide variety of industrial fields. The classification of proteases and the multiple sources used to isolate alkaline protease-producing microbes are summarized in this paper. It has been discussed how different physicochemical factors affect alkaline proteases. The paper also briefly discusses the ideal pH and temperature for alkaline protease-producing microorganisms and industrial applications

Key Words: Alkaline protease, hydrolases, physicochemical parameters.

1. INTRODUCTION:

Chemicals are globular proteins and, as different proteins, are made out of lengthy chains of amino acids that crease into three-layered items. Every novel amino corrosive grouping makes a particular design with exceptional properties. Compounds, profoundly specific biocatalysts, have been utilized in the food business for many years and furthermore assume a significant part in different enterprises like cleansers, drugs, mash and paper, material assembling. Chemicals can separate specific mixtures. Atoms on which chemicals act are called substrates and are changed over into items. Amylases, which convert starch into straightforward sugars, proteases complete separate of proteins, cellulases, do separate of cellulose, and lipases, what split lipids into glycerol and unsaturated fats, are the absolute generally common. (Mojsov K, 2012).

2. Classification of chemicals:

The EC order framework is partitioned into seven classifications of fundamental capability: EC1 Oxidoreductases, EC2 Transferases, EC3 Hydrolases, EC4 Lyases, EC5 Isomerases, EC6 Ligases and EC7 Translocases, (McDonald, 2009).



Table 2.1: The EC System of Classification of Peptidases

Sub-subclass	Peptidase type	Number of entries
3.4.11	Aminopeptidases	26
3.4.13	Dipeptidases	23
3.4.14	Dipeptidyl-peptidases and tripeptidyl-peptidases	14
3.4.15	Peptidyl-dipeptidases	6
3.4.16	Serine-type carboxypeptidases	6
3.4.17	<u>Metallo</u> carboxypeptidases	25
3.4.18	Cysteine-type carboxypeptidases	1
3.4.19	Omega peptidases	16
3.4.21	Serine endopeptidases	122
3.4.22	Cysteine endopeptidases	71
3.4.23	Aspartic endopeptidases	52

2.1 Classification of proteases:

Proteases are a large group of hydrolases that catalyze the hydrolysis of proteins by cleaving peptide bonds between amino acid residues of other proteins. (Shankar et al., 2011) Proteases (EC 3.4.21-24 and 99; peptidyl peptide hydrolases) hydrolyze proteins through the addition of water to peptide bonds and catalyze peptide synthesis in organic and low-moisture solvents (Beg et al., 2003).

Proteases are members of the hydrolase class of enzymes and catalyze the typical hydrolysis of peptide bonds; nevertheless, proteases do not readily fit into the established classification scheme for enzymes. Currently, there are three basic criteria used to categorise proteases, type of catalyzed reaction, chemical make-up of catalytic site and structural relationship. (Barret, 1994)

2.2 Type of reaction catalyzed:

These enzymes can be broadly divided into two groups depending on where they act, exopeptidases, which break C- or N-terminal peptide bonds, and endopeptidases, which cleave interior peptide bonds. Endopeptidases set themselves apart by acting preferentially on peptide bonds in the inside, away from the N or C termini, of polypeptide chains. (Sharma et al., 2019)

2.3 Chemical nature of catalytic site

Aspartic proteases, Serine proteases, Metallo proteases, Cysteine proteases, Threonine proteases, Glutamic proteases, Serine proteases, and Mixed proteases are the seven subgroups based on their catalytic mechanism (MEROPS, n.d.). Alkaline proteases are proteases that are active in the neutral to alkaline pH range (EC.3.4.21-24, 99). They either have a metallopeptidase or a serine centre these are metalloprotease and serine protease respectively. They are the most significant group of enzymes utilized in industry. (Gupta et al., 2002b).

Table 2.1.2: Protease types according to catalytic site

Type	Examples
Aspartic	pepsin A, nepenthesin
Cysteine	papain, bleomycin hydrolase
Glutamic	scytalidoglutamic peptidase
Metallo	Thermolysin, collagenase V
Mixed	polycystin-1
Serine	chymotrypsin A, prolyl oligopeptidase
Threonine	archaeal proteasome
Unknown	yabG protein, collagenase



On the basis of the chemistry of the catalytic site and an unknown catalytic type, seven types of proteases are recognized. The Web pages of the MEROPS database contain additional details about the example of enzymes listed above. (MEROPS, n.d. 1) (Refer Internet Resources) https://www.ebi.ac.uk/merops/cgi-bin/family_index?type=P

4. Structural Relationship

Proteases are categorised in the MEROPS system using structural characteristics that are thought to represent evolutionary links. Based on similarities in the amino acid sequence, closely related peptidases are grouped into families. Families that show signs of a common ancestor come together to establish clans, (MEROPS, n.d. 2). https://www.ebi.ac.uk/merops/cgi-bin/clan_index?type=P

5. Sources of Proteases:

Until the mid 1970s, plant and creature sources were viewed as the best wellsprings of compounds. Today, in any case, microbial catalysts are turning out to be progressively significant because of their specialized and monetary benefits, (Mohamed et al., 2007). The primary wellsprings of compounds were creatures, (for example, calf stomach), plants (like pineapples, figs, and papaya), and microorganisms (like *Bacillus* and *Pseudomonas*). In any case, the development of proteins from plant and creature sources is limited for climatic and moral reasons, (Shafee et al. 2005, Rao et al., 1998).

Microorganisms are a positive wellspring of proteases as they might be created in high amounts in a short measure of time utilizing standard maturation methods and proposition a copious and reliable stock of the ideal item. Moreover, microbial proteins are durable and can be saved for quite some time in under ideal conditions without experiencing a significant reduction in action. (Gupta et al., 2002a).

As per whether they are dynamic in unbiased, acidic, or soluble conditions, microbial proteases are classified into three classes. The calfskin, cleanser, food, and material areas are among those that expect to utilize soluble proteases widely, (Rathod et al., 2013). Proteolytic proteins that function admirably at soluble pH are alluded to as antacid proteases, (Barret, 1994; Gupta et al., 2002). These proteins can be created by microorganisms both intracellularly and extracellularly. Protease seclusion from organisms is simple and reasonable, particularly for extracellular proteases. Creation of chemicals by organisms is ordinarily unaffected via occasional variances in the accessibility of unrefined components, (Roja et al., 2012).

Antacid protease is for the most part created by microbes, with the sort *Bacillus* being the fundamental maker, (Gupta et al., 2002b, Kalisz HM (1988), Kumar and Takagi (1999), and Rao et al., 1998). *Pseudomonas* sp. is one more kind of bacterium that has been recognized as a possible maker, (Bayoudh et al., 2000, Ogino et al., 1999). Actinomycetes strains are the most wanted hotspot for soluble proteases, (Petinate et al., 1999) and *Aspergillus* is the most broadly utilized class of parasites, (Chakrabarti et al. 2000, Rajamani et al., 1987). *Rhizopus* (Banerjee and Bhattacharyya (1993) and *Conidioborus* (Bhosale et al., 1995) are two different genera that likewise yield antacid proteases. Yeasts incorporate *Candida*, broadly researched as a possible maker of basic protease, (Poza et al., 2001.).

6. Isolation sources for alkaline protease producing microbes:

For confinement of soluble protease delivering organisms different sources have been accounted for. Ibrahim et al gathered Dregs and water tests from hyper saline sodalakes in northern Egypt and separated basic protease delivering alkalophilic microorganisms. By utilizing Horikoshi-I soluble medium with some adjustment, (Ibrahim et al., 2015). Gasp et al gathered Soil tests from side streets in Chennai. Completed sequential weakenings of the, spread on supplement agar plates and brooded for 24 h at 37°C, (Gasp et al., 2015). Roja et al. detached alkalophilic microscopic organisms from basic soil tests got from dark cotton soil, groundnut ranches, milk handling units, and the Kotappakonda slope area of Chittur Region, A.P. The microorganisms were screened on a skim milk agar plate, (Roja et al., 2012). In waterfront Gujarat, India, Dodia et al confined bacterial strains that produces antacid protease in regular hyper-saline climate, (Dodia et al., 2006).

Ibrahim et al chose the settlements which shows zone of leeway around it because of casein hydrolysis, (Ibrahim et al., 2015). Roja et al distinguished alkalophilic microscopic organisms delivering protease by the skim milk hydrolysis around settlements, (Roja et al., 2012).

7. Effect of physico-substance boundary on protease creation

Ibrahim et al detailed that different carbon sources distinctively affect the development of protease from *Bacillus* sp. NPST-AK15. Among the different carbon sources, fructose showed expanded protease creation while glucose or maltose prompted an uncommon diminishing in the chemical creation. Among the nitrogen source greatest



yield got in medium containing yeast remove, trailed by skim milk, gelatin, casein, and other natural nitrogen sources, separately. Air circulation showed influence on bacterial development and chemical creation, with expansion in air circulation of up to 200 rpm brought about an ascent in development and protein creation, (Ibrahim et al., 2015).

8. Purging of Protease:

Aikat et al to some degree cleansed the protease from *Rhizopus oryzae* by charcoal and detailed 85% compound recuperation, affirmed incomplete sanitization by local PAGE, (Aikat et al., 2000). Gessesse et al filtered the protein by precipitation of culture filtrate utilizing strong ammonium sulfate, (Gessesse et al., 1997). Gasp et al completed precipitation of rough compound planning with ammonium sulfate, separated the reaped culture through Whatman no. 1 filter paper. centrifuged the filtrate, ammonium sulfate was added to the sans cell culture filtrate. The encouraged protein was isolated by centrifugation and dialysed, (Gasp et al., 2015). El-Shanshoury et al cleansed protease by precipitation in chilled ethanol and in this way gel filtration on Sephadex G-120 and cboxymethyl Sepharose, (El-Shanshoury et al 1995).

9. Application of Proteases :

In 2021 the worldwide protein market was US\$11.47 billion and is supposed to rise US\$ 20.31 billion out of 2030 at a build yearly pace of 6.5% per annum. (Compounds Market Size, Offer and Patterns Examination Report, 2030. (n.d.)). Proteases are one of the main gatherings of modern catalysts, representing more than 65% of the all out modern compound market (Sundararajan et al., 2011, Annamalai et al., 2014, Zanthorlin et al., 2010). Additionally, microbial proteases represent around 40% of the worldwide creation of catalysts (Haddar et al., 2009, Raval et al., 2014). Microbial antacid proteases have various applications in different modern fields, like facilities, food, cleansers, and calfskin, (Sharma et al., 2019). Proteins are presented by business catalyst makers for extensive variety of uses. The top areas using around 75% of mechanically fabricated proteins are cleanser (37%), material (12%), starch (11%), pastry shops (8%) and creature feed (6%), (Singh et al., 2011). Before long; microbial proteins might supplant catalysts from different sources and presently represent almost 90% of the complete market. This is on the grounds that microbial cells are incredible frameworks for compound creation. Subsequently, there is extraordinary stimulus for enormous scope research exercises on recombinant proteins. There is a ton of use for basic proteases in the cleanser, cowhide tanning, and food areas (Kelly et al., 1976, Godfrey et al., 1985) and furthermore in the creature feed industry, peptide blend, modern family garbage removal, the visual business, clinical applications, silk degumming, (Rao et al., 1998), materials and waste water treatment (Velooralappil et al., 2013).

In the food processing sector, proteases are extensively used. They are frequently employed in the preparation of dairy products, baked goods, meat tenderization, and processing of animal proteins, vegetable proteins, seafood, and bioactive peptides. Enhancing food's functional characteristics, nutrition, digestibility, flavour quality, taste, and visibility, increasing antioxidant capacity, and preventing food poisoning and negative effects are the main goals of utilising enzymes in the food industry, (Varia et al., 2019). The baking process benefits from the employment of microbial proteases because they can degrade the complex network of glutenins and gliadins. Baking is enhanced by microbial proteases. Proteases are added to dough to enhance volume, improve handling characteristics, and make soft bread with a good crumble and crusty structure. It has been observed that the production of softer dough using proteases is due to the gluten hydrolysis that occurs in the dough (Varia et al., 2019).

REFERENCES:

1. Aikat, K., Maiti, T. K., & Bhattacharyya, B. C. (2001). Decolorization and purification of crude protease from *Rhizopus oryzae* by activated charcoal and its electrophoretic analysis. *Biotechnology letters*, 23(4), 295-301.
2. Annamalai, N., Rajeswari, M. V., & Balasubramanian, T. (2014). Extraction, purification and application of thermostable and halostable alkaline protease from *Bacillus alveayuensis* CAS 5 using marine wastes. *Food and Bioproducts Processing*, 92(4), 335-342.
3. Banerjee, R., & Bhattacharyya, B. C. (1993). Kinetic properties of extracellular alkaline protease of *Rhizopus oryzae*. *Journal of fermentation and bioengineering*, 75(5), 380-382.
4. Barret, A. J. (1994). Proteolytic enzymes: serine and cysteine peptidases. *Methods Enzymol.*, 244, 1-765.
5. Bayoudh, A., Gharsallah, N., Chamkha, M., Dhouib, A., Ammar, S., & Nasri, M. (2000). Purification and characterization of an alkaline protease from *Pseudomonas aeruginosa* MN1. *Journal of Industrial Microbiology and Biotechnology*, 24(4), 291-295.
6. Beg, Q. K., Sahai, V., & Gupta, R. (2003). Statistical media optimization and alkaline protease production from *Bacillus mojavensis* in a bioreactor. *Process Biochemistry*, 39(2), 203-209.



7. Bhosale, S. H., Rao, M. B., Deshpande, V. V., & Srinivasan, M. C. (1995). Thermostability of high-activity alkaline protease from *Conidiobolus coronatus* (NCL 86.8. 20). *Enzyme and Microbial Technology*, 17(2), 136-139.
8. Chakrabarti, S. K., Matsumura, N., & Ranu, R. S. (2000). Purification and characterization of an extracellular alkaline serine protease from *Aspergillus terreus* (IJIRA 6.2). *Current Microbiology*, 40(4), 239-244.
9. Dodia, M. S., Joshi, R. H., Patel, R. K., & Singh, S. P. (2006). Characterization and stability of extracellular alkaline proteases from halophilic and alkaliphilic bacteria isolated from saline habitat of coastal Gujarat, India. *Brazilian Journal of Microbiology*, 37(3), 276-282.
10. El-Shanshoury, A. E. R. R., El-Sayed, M. A., Sammour, R. H., & El-Shouny, W. A. (1995). Purification and partial characterization of two extracellular alkaline proteases from *Streptomyces corchorusii* ST36. *Canadian journal of microbiology*, 41(1), 99-104.
11. Enzymes Market Size, Share & Trends Analysis Report, 2030. (n.d.). Retrieved September 25, 2022, from [https://www.grandviewresearch.com/industry-analysis/enzymes-industry+](https://www.grandviewresearch.com/industry-analysis/enzymes-industry)
12. Gessesse, A., & Gashe, B. A. (1997). Production of alkaline protease by an alkaliphilic bacteria isolated from an alkaline soda lake. *Biotechnology letters*, 19(5), 479-481.
13. Godfrey, T., & Reichelt, J. (1982). *Industrial enzymology: the application of enzymes in industry*.
14. Gupta, R., Beg, Q., & Lorenz, P. (2002b). Bacterial alkaline proteases: molecular approaches and industrial applications. *Applied microbiology and biotechnology*, 59(1), 15-32.
15. Gupta, R., Beg, Q., Khan, S., & Chauhan, B. (2002a). An overview on fermentation, downstream processing and properties of microbial alkaline proteases. *Applied microbiology and biotechnology*, 60(4), 381-395.
16. Haddar, A., Bougatef, A., Agrebi, R., Sellami-Kamoun, A., & Nasri, M. (2009). A novel surfactant-stable alkaline serine-protease from a newly isolated *Bacillus mojavensis* A21. *Purification and characterization*. *Process Biochemistry*, 44(1), 29-35.
17. Ibrahim, A. S., Al-Salamah, A. A., Elbadawi, Y. B., El-Tayeb, M. A., & Ibrahim, S. S. S. (2015). Production of extracellular alkaline protease by new halotolerant alkaliphilic *Bacillus* sp. NPST-AK15 isolated from hyper saline soda lakes. *Electronic Journal of Biotechnology*, 18(3), 236-243.
18. Kalisz, H. M. (1988). Microbial proteinases. *Enzyme studies*, 1-65.
19. Kelly, C. T., & Fogarty, W. M. (1976). Microbial alkaline enzymes. *Process biochemistry*, 11, 3-9.
20. Kumar, C. G., & Takagi, H. (1999). Microbial alkaline proteases: from a bioindustrial viewpoint. *Biotechnology advances*, 17(7), 561-594.
21. McDonald, A. (2009). ExplorEnz- the enzyme database. <https://www.enzyme-database.org>. Retrieved September 25, 2022, from <https://www.enzyme-database.org/class.php>
22. MEROPS. (n.d. 1). Retrieved September 25, 2022, from https://www.ebi.ac.uk/merops/cgi-bin/family_index?type=P
23. MEROPS. (n.d. 2). Retrieved September 25, 2022, from https://www.ebi.ac.uk/merops/cgi-bin/clan_index?type=P
24. Mohamed, L., Zakaria, M., Ali, A., Senhaji, W., Mohamed, O., Mohamed, E., ... & Mohamed, J. (2007). Optimization of growth and extracellular glucoamylase production by *Candida famata* isolate. *African Journal of Biotechnology*, 6(22).
25. Mojsov, K. (2012). Microbial alpha-amylases and their industrial applications: a review. *International Journal of Management, IT and Engineering (IJMIE)*, 2(10), 583-609.
26. Ogino, H., Watanabe, F., Yamada, M., Nakagawa, S., Hirose, T., Noguchi, A., ... & Ishikawa, H. (1999). Purification and characterization of organic solvent-stable protease from organic solvent-tolerant *Pseudomonas aeruginosa* PST-01. *Journal of bioscience and bioengineering*, 87(1), 61-68.
27. Pant, G., Prakash, A., Pavani, J. V. P., Bera, S., Deviram, G. V. N. S., Kumar, A., ... & Prasuna, R. G. (2015). Production, optimization and partial purification of protease from *Bacillus subtilis*. *Journal of Taibah University for Science*, 9(1), 50-55.
28. Petinate, S. D. G., Branquinha, M. H., Coelho, R. R. R., And, A. V., & Giovanni-De-Simone, S. (1999). Purification and partial characterization of an extracellular serine-proteinase of *Streptomyces cyaneus* isolated from Brazilian cerrado soil. *Journal of applied microbiology*, 87(4), 557-563.
29. Poza, M., De Miguel, T., Sieiro, C., & Villa, T. G. (2001). Characterization of a broad pH range protease of *Candida caseinolytica*. *Journal of Applied Microbiology*, 91(5), 916-921.
30. Rajamani, S., & Hilda, A. (1987). Plate assay to screen fungi for proteolytic activity. *Current Science*, 56(22), 1179-1181.



31. Rao, M. B., Tanksale, A. M., Ghatge, M. S., & Deshpande, V. V. (1998). Molecular and biotechnological aspects of microbial proteases. *Microbiology and molecular biology reviews*, 62(3), 597-635.
32. Rathod, M. G., & Pathak, A. P. (2013). Production dynamics of extracellular alkaline protease by *Bacillus globisporus* LAP19: a Lonar soda lake isolate. *Conservation of medicinal plants and their utilization* (1), 46-53.
33. Raval, V. H., Pillai, S., Rawal, C. M., & Singh, S. P. (2014). Biochemical and structural characterization of a detergent-stable serine alkaline protease from seawater haloalkaliphilic bacteria. *Process Biochemistry*, 49(6), 955-962.
34. Roja Rani, M., Lalitha Kumara, B., & Siva Prasad, D. (2012). Isolation and screening of alkaline protease producing bacteria and induction of overproducing *Bacillus licheniformis* mutants through UV irradiation. *IOSR J. Pharmacy*, 1, 1-14.
35. Shafee, N., Aris, S. N., Rahman, R. N. Z. A., Basri, M., & Salleh, A. B. (2005). Optimization of environmental and nutritional conditions for the production of alkaline protease by a newly isolated bacterium *Bacillus cereus* strain 146. *J Appl Sci Res*, 1(1), 1-8.
36. Shankar, S., Rao, M., & Laxman, R. S. (2011). Purification and characterization of an alkaline protease by a new strain of *Beauveria* sp. *Process biochemistry*, 46(2), 579-585.
37. Sharma, M., Gat, Y., Arya, S., Kumar, V., Panghal, A., & Kumar, A. (2019). A review on microbial alkaline protease: an essential tool for various industrial approaches. *Industrial Biotechnology*, 15(2), 69-78.
38. Singh SURENDRA, Sharma, VINNI, Soni, M. L., & Das, SHIPRA (2011). Biotechnological applications of industrially important amylase enzyme. *International Journal of Pharma and Bio Sciences*, 2(1), 486-496.
39. Sundararajan, S., Kannan, C. N., & Chittibabu, S. (2011). Alkaline protease from *Bacillus cereus* VITSN04: potential application as a dehairing agent. *Journal of bioscience and bioengineering*, 111(2), 128-133.
40. Varia, A. D., Shukla, V. Y., & Tipre, D. R. (2019). ALKALINE PROTEASE-A VERSATILE ENZYME.
41. Velloorvalappil, N. J., Robinson, B. S., Selvanesan, P., Sasidharan, S., Kizhakkepawothail, N. U., Sreedharan, S., & Sailas, B. (2013). Versatility of microbial proteases. *Advances in enzyme research*, 2013.
42. Zanthorlin, L. M., Facchini, F. D. A., Vasconcelos, F., Bonugli-Santos, R. C., Rodrigues, A., Sette, L. D., & Bonilla-Rodriguez, G. O. (2010). Production, partial characterization, and immobilization in alginate beads of an alkaline protease from a new thermophilic fungus *Myceliophthora* sp. *The Journal of Microbiology*, 48(3), 331-336.